

# Antibacterial activity of garlic extract on streptomycin-resistant *Staphylococcus aureus* and *Escherichia coli* solely and in synergism with streptomycin

M. N. Palaksha,  
Mansoor Ahmed<sup>1</sup>,  
Sanjoy Das

Aditya Institute of Pharmaceutical Sciences and Research, A.D.B. Road, Aditya Nagar, Surampalem, East Godavari District, Andhra Pradesh, <sup>1</sup>Department of Pharmacology, Sri Siddaganga College of Pharmacy, B. H. Road, Tumkur, Karnataka, India

## Address for correspondence:

Mr. M. N. Palaksha, Aditya Institute of Pharmaceutical Sciences and Research, A. D. B. Road, Aditya Nagar, Surampalem, East Godavari District, Andhra Pradesh – 533 437, India.

DOI: 10.4103/0976-9668.71666

## Abstract

This study focuses the significant antibacterial activity of Garlic (*Allium sativum* Linn.) extract on streptomycin-resistant strains solely and in synergism with streptomycin. Gram-positive *Staphylococcus aureus* ATCC BAA 1026 and gram-negative *Escherichia coli* ATCC 10536 were made resistant to standard antibiotic streptomycin used as a control in the experiment. Zones of inhibition of different treatment groups were measured by agar-well-diffusion assay and compared with control. Statistical comparison of sole extract and streptomycin synergism with streptomycin control had proved it significant.

**Key words:** Antibacterial, antibiotic, *Escherichia coli*, resistance, *Staphylococcus aureus*, synergism

## INTRODUCTION

Since ancient time, naturally occurring plants have played an important role in the discovery of new therapeutic agents.<sup>[1]</sup> Almost all antibiotics are subjected to the problem of bacterial resistance. Therefore, newer herbal antibacterial compounds from plants and their semisynthetic derivatives to overcome the resistance are under investigation.

Garlic (*Allium sativum* Linn.) has an important dietary and medicinal role for centuries. Its therapeutic uses include beneficial effects on the cardiovascular system, antibiotic, anticancer, anti-inflammatory, hypoglycemic, and hormone-like effects.<sup>[2]</sup> Garlic extracts have been used to treat infections for thousands of years.<sup>[3]</sup> Its typical pungent odor and antibacterial activity depend on allicin, which is produced by enzymatic (alliin lyase) hydrolysis of alliin after cutting and crushing of the cloves.<sup>[4]</sup>

Streptomycin was widely used for more than four decades, which is still effective against gram-positive and gram-

negative bacteria. This study reveals better efficacy of garlic extract and its streptomycin synergism than streptomycin on resistant strains.

## MATERIALS AND METHODS

### Source of bacterial strains

Both gram-positive *Staphylococcus aureus* ATCC BAA 1026 and gram-negative *Escherichia coli* ATCC 10536 were collected from clinical specimens of Padmashree Diagnostic Centre, Tumkur, India.

### Development of streptomycin resistance in selected bacterial strains

Each organism was subcultured from a nutrient agar (Qualigens Fine Chemicals, Mumbai, India) slant to standard methods broth (Human Diagnostic and Surgichem, kolkota, India), PH 7.8, and incubated overnight. With stock solutions of standard antibiotic streptomycin (gift sample on request from Karnataka Antibiotics and Pharmaceuticals Limited, Bangalore, India), which were prepared by diluting weighed aliquots of this drug in

sterile 1% phosphate buffer PH 6.0, twofold dilutions were prepared daily. The dilution series were usually consisted of ten 100× 13 mm test tubes each containing 0.5 ml of the antibiotic dilution. To each tube was added 1.5 ml of a 1:100 dilution in broth of the 18–24 h broth culture prepared above, and all the tubes were incubated at 37°C for 24 h. The last tube showing inhibition of the organism in the dilution series indicated the initial sensitivity of the strain in micrograms of the antibiotic. The second tube showing growth in dilution series was selected for preparing 1:100 broth dilutions for the second exposure to streptomycin dilution series. To increase the resistance of the strain to the particular antibiotic, the procedure described above was repeated.<sup>[5]</sup>

### Preparation of aqueous garlic extract

Fresh garlic (*Allium sativum* L.) bulbs were purchased from local market. The bulbs were peeled, weighed (100 gm) and cleaned. Cleaned cloves were surface-sterilized by immersing them into 70% (v/v) ethanol for 60s.<sup>[6]</sup> Residual ethanol on surface was evaporated in sterile laminar airflow chamber followed by homogenizing aseptically in sterile mortar and pestle. The homogenized mixture was filtered through sterile cheesecloth. This extract was considered as the 100% concentration of the extract. The concentrated mother extract was further diluted to 75% and 50% by mixing with appropriate sterile distilled water.<sup>[7]</sup>

### Testing of antibacterial activity using agar well diffusion method

Resistant bacterial strains were inoculated into 10 ml of sterile nutrient broth, and incubated at 37 °C for 8 h. Each culture was swabbed on the surface of sterile nutrient agar plate in duplicate. In each agar plate of both sets, five wells were prepared with the help of sterilized cork borer of 10 mm diameter. In the wells of first plate of each set, 100 µl test samples of following concentrations: (1) standard streptomycin 10 mg/ml in sterile distilled water; (2) 50% sterile garlic extract; (3) streptomycin 10 mg/ml in 50% sterile garlic extract; (4) streptomycin 10 mg/ml in 75% sterile garlic extract; (5) streptomycin 10 mg/ml in 100% sterile garlic extract) were added by using micropipette. In the wells of second plate of each set, 100 µl test samples of the following concentrations: (1) standard streptomycin 10 mg/ml in sterile distilled water; (2) 50% sterile garlic extract; (3) streptomycin 10 mg/ml in 50% sterile garlic extract; (4) streptomycin 15 mg/ml in 50% sterile garlic extract; (5) streptomycin 20 mg/ml in 50% sterile garlic extract) were added. Every plate used according to the aforementioned procedure was performed in triplicate for statistical average.

## RESULTS

Mean zones of inhibition were expressed in mm ± standard error of mean. Mean zones of inhibition of

different treatment groups were measured by agar-well-diffusion assay and compared with the control. Statistical comparison of sole garlic extract and streptomycin synergism (same concentration of streptomycin in garlic extract of different strengths and different concentration of streptomycin in the garlic extract of same strength as stated in Tables 1 and 2, respectively) with streptomycin control by one-way ANOVA post-test using the software graphpad Instat 3 (trial) had proved it significant. Figures 1 and 2 illustrate Table 1 whereas Figures 3 and 4 do the same for Table 2.

## DISCUSSION

The findings of this study reveal the distinct antibacterial profile of *Allium sativum* Linn. solely and in streptomycin synergism against streptomycin-resistant *S. aureus* ATCC BAA 1026 and *E. coli* ATCC 10536 as witnessed from prominent zones of inhibition. *E. coli* is a common pathogenic bacteria for urinary tract infection and *S. aureus* is the cause of pneumonia and several infections in gut, urinary tract, etc. Use of garlic extract solely is fruitful.

**Table 1: Inhibition of resistant bacteria due to sole garlic extract and synergism of same concentration of streptomycin in garlic extract of different strengths in the presence of streptomycin control**

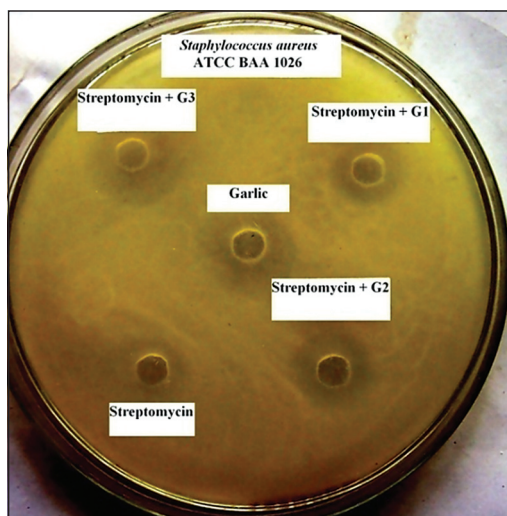
Drug	Dose	Zone of inhibition (mm ± S.E.M.)	
		<i>S. aureus</i> ATCC BAA 1026	<i>E. coli</i> ATCC 10536
Streptomycin <sup>a</sup> (control)	10 mg/ml	7 ± 0.2887	8 ± 0.2887
Sterile garlic extract <sup>a</sup>	50%	14 ± 0.5775**	14.5 ± 0.2887**
Streptomycin <sup>b</sup>	10 mg/ml	15 ± 0.2887**	16 ± 0.2887**
Streptomycin <sup>c</sup>	10 mg/ml	22 ± 0.2887**	24 ± 0.5774**
Streptomycin <sup>d</sup>	10 mg/ml	26 ± 0.2887**	28 ± 0.2887**

<sup>a</sup>Solvent: sterile distilled water. <sup>b</sup>Solvent: 50% sterile garlic extract. <sup>c</sup>Solvent: 75% sterile garlic extract. <sup>d</sup>Solvent: 100% sterile garlic extract. \*\**P* < 0.01 as compared with control according to one-way ANOVA post-test.

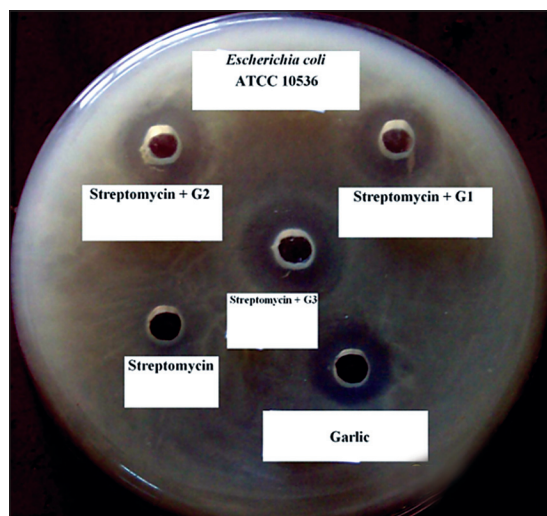
**Table 2: Inhibition of resistant bacteria due to sole garlic extract and synergism of different concentration of streptomycin in garlic extract of same strength in presence of streptomycin control**

Drug	Dose	Zone of inhibition (mm ± S.E.M.)	
		<i>S. aureus</i> ATCC BAA 1026	<i>E. coli</i> ATCC 10536
Streptomycin <sup>a</sup> (control)	10 mg/ml	6 ± 0.2887	7 ± 0.2887
Garlic extract <sup>a</sup>	50%	15 ± 0.2887**	14 ± 0.2887**
Streptomycin <sup>b</sup>	10 mg/ml	16 ± 0.2887**	15 ± 0.2887**
Streptomycin <sup>b</sup>	15 mg/ml	19 ± 0.5774**	17 ± 0.2887**
Streptomycin <sup>b</sup>	20 mg/ml	20 ± 0.2887**	19 ± 0.5774**

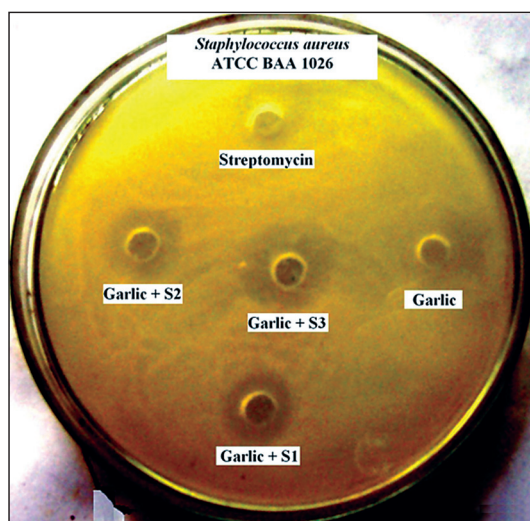
<sup>a</sup>Solvent: sterile distilled water. <sup>b</sup>Solvent: 50% sterile garlic extract. \*\**P* < 0.01 as compared with control according to one-way ANOVA post-test.



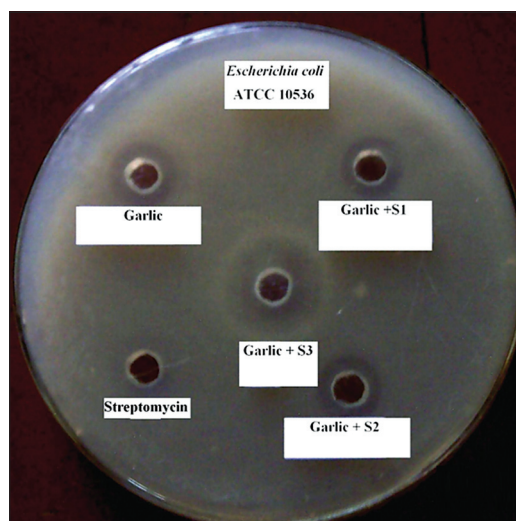
**Figure 1:** G1: 50% garlic extract; G2: 75% garlic extract; G3: 100% garlic extract. The concentration of streptomycin alone and in G1, G2, G3, etc. is 10 mg/ml. Concentration of garlic used alone is 50%.



**Figure 2:** G1: 50% garlic extract; G2: 75% garlic extract; G3: 100% garlic extract. The concentration of streptomycin alone and in G1, G2, G3, etc. is 10 mg/ml. Concentration of garlic used alone is 50%.



**Figure 3:** S1: 10 mg/ml; S2: 15 mg/ml; S3: 20 mg/ml of streptomycin. The concentration of garlic extract used alone and with S1, S2, S3, etc. is 50% garlic extract. Concentration of streptomycin used alone is 10 mg/ml.



**Figure 4:** S1: 10 mg/ml; S2: 15 mg/ml; S3: 20 mg/ml of streptomycin. The concentration of garlic extract used alone and with S1, S2, S3, etc. is 50% garlic extract. Concentration of streptomycin used alone is 10 mg/ml.

Synergistic use can prevent the pathogenic organism grow their resistance against antibiotic.

## ACKNOWLEDGMENTS

The author (M.N.P.) would like to convey his sincere gratitude to his colleague Mr. C. Pallavan for his kind support and necessary help to make the original research paper. He is thankful to his former principal Dr. D. Banji for his advice and valuable suggestions. Finally, he would like to thank all laboratory staffs of Sri Siddaganga College of Pharmacy, B.H. Road, Tumkur, Karnataka for their continuous help regarding this work.

## REFERENCES

1. Mohanty JP, Nath LK, Bhuyan NR, Mahapatra SK. Antibacterial spectrum of *Kaempferia rotunda* Linn. and *Eupatorium cannabinum* Linn. *Adv Pharmacol Toxicol* 2008;9:45-50.
2. Jonkers D, van den Broek E, van Dooren I, Thijs C, Dorant E, Hageman G, *et al.* Antibacterial effect of garlic and omeprazole on *Helicobacter pylori*. *J Antimicrob Chemother* 1999;43:837-9.
3. Hahn G. Garlic: The science and therapeutic application of *Allium sativum* Linn. and related species. In: Koch HP, Lawson LD, editors. 2<sup>nd</sup> ed. Baltimore: Williams and Wilkins; 1996. p. 1-24.
4. Ellmore GS, Feldberg RS. Alliin lyase localization in bundle sheaths of Garlic cloves (*Allium sativum* Linn.) *Am J Bot* 1994;81:89-94.
5. Price CW, Randall WA, Chandler VL, Reedy RJ. Observation on the *in vivo* and *in vitro* development of bacterial resistance to streptomycin.

- J Bacteriol 1947;53:481-8.
6. Kalyan KD. An introduction to plant tissue culture. 1<sup>st</sup> ed. Calcutta: New Central Book Agency (P) Ltd; 2000. p. 37-9.
  7. Durairaj S, Sangeetha S, Lakshmanaperumalsamy P. *In vitro*

antibacterial activity and stability of garlic extract at different pH and temperature. Electron J Biol 2009;5:5-10.

**Source of Support:** Nil, **Conflict of Interest:** None declared.

## Author Help: Online submission of the manuscripts

Articles can be submitted online from <http://www.journalonweb.com>. For online submission, the articles should be prepared in two files (first page file and article file). Images should be submitted separately.

### 1) **First Page File:**

Prepare the title page, covering letter, acknowledgement etc. using a word processor program. All information related to your identity should be included here. Use text/rtf/doc/pdf files. Do not zip the files.

### 2) **Article File:**

The main text of the article, beginning with the Abstract to References (including tables) should be in this file. Do not include any information (such as acknowledgement, your names in page headers etc.) in this file. Use text/rtf/doc/pdf files. Do not zip the files. Limit the file size to 1024 kb. Do not incorporate images in the file. If file size is large, graphs can be submitted separately as images, without their being incorporated in the article file. This will reduce the size of the file.

### 3) **Images:**

Submit good quality color images. Each image should be less than **2048 kb (2 MB)** in size. The size of the image can be reduced by decreasing the actual height and width of the images (keep up to about 6 inches and up to about 1800 x 1200 pixels). JPEG is the most suitable file format. The image quality should be good enough to judge the scientific value of the image. For the purpose of printing, always retain a good quality, high resolution image. This high resolution image should be sent to the editorial office at the time of sending a revised article.

### 4) **Legends:**

Legends for the figures/images should be included at the end of the article file.